

Review

Open Access

Ivermectin: does P-glycoprotein play a role in neurotoxicity?

Geoffrey Edwards*^{1,2}

Address: ¹Department of Pharmacology and Therapeutics, The University of Liverpool, Sherrington Buildings, Ashton Street, Liverpool, UK and ²Division of Molecular and Biochemical Parasitology, Liverpool School of Tropical Medicine, Pembroke Place, Liverpool, UK

Email: Geoffrey Edwards* - ge1000@liverpool.ac.uk

* Corresponding author

from Report of a Scientific Working Group on Serious Adverse Events following Mectizan® treatment of onchocerciasis in *Loa loa* endemic areas Shrigley Hall Hotel, Manchester, UK, 28 – 30 May 2002

Published: 24 October 2003

Filaria Journal 2003, 2(Suppl 1):S8

This article is available from: <http://filariajournal.com/content/2/S1/S8>

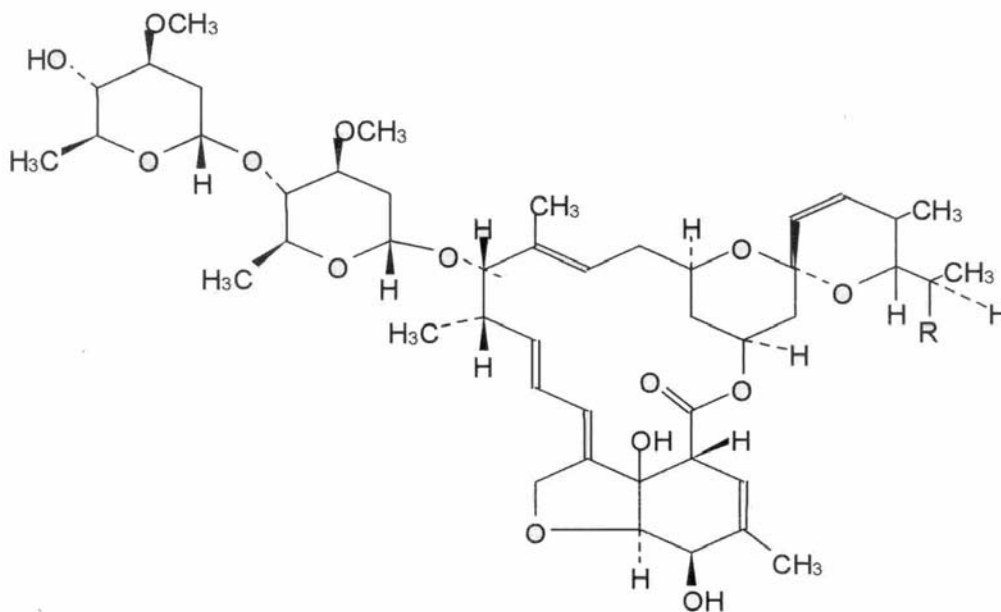
Abstract

The macrocyclic lactone ivermectin (Mectizan®) is widely used for the control of human filarial infections, particularly as a donated product for onchocerciasis and lymphatic filariasis. In the case of control of lymphatic filariasis in Africa, it is used in combination with donated albendazole. In areas co-endemic for Onchocerciasis and *Loa loa*, serious adverse reactions have been observed in patients with apparently high microfilaria counts of *Loa loa*. Recent findings suggest that the severe central nervous system side effects seen in various vertebrates following ivermectin treatment may be due to an absence of, or functional deficiency in P-glycoprotein. P-glycoprotein is expressed in the apical membrane of brain capillary epithelial cells and is responsible for limiting the brain penetration of a range of compounds. Toxicity of ivermectin in some collie dogs may be explained by a 4-bp deletion mutation of the *mdr1* gene resulting in a frame shift, generating stop codons that prematurely terminate synthesis of P-glycoprotein. Additionally, sub-populations of CF-1 identified as expressing reduced levels of P-glycoprotein exhibit increased toxicity to substrates of this transporter. Furthermore, while the traditional view of drug-drug interactions is alteration in drug clearance mediated through a change in hepatic drug metabolism, some of these changes may arise through competition for binding sites on P-glycoprotein in the blood-brain barrier, resulting in reduced extracellular efflux and enhanced CNS toxicity. In conclusion, P-glycoprotein is an integral component of the human blood brain barrier and plays a central role in limiting drug uptake into the brain. Altered expression or function of p-glycoprotein could conceivably allow elevation of brain concentrations of ivermectin and produce severe neurotoxicity. This might arise through a genetic polymorphism in p-glycoprotein or co-administration of ivermectin with a drug or foodstuff that might inhibit this efflux transporter.

Background

Although clinical and veterinary usage of the broad-spectrum anthelmintic agent ivermectin has generally been free of serious adverse events since 1991, a number of clinical cases with and without neurological manifestations, including coma lasting 2–3 days, have been reported after ivermectin treatment of individuals infected

with *Onchocerca volvulus* who also had concomitant *Loa loa* infection with a high levels of microfilariae, typically > 30,000 microfilariae /ml of blood [1,2]. Although a cause and effect relationship has yet to be established, it is advised that individuals who warrant treatment with ivermectin for any reason and have had significant exposure to *Loa loa*-endemic areas of West and Central Africa, pre-

**Figure 1**

Chemical structure of ivermectin. Ivermectin contains not less than 93.0% of 22,23 dihydroavermectin B_{1a} (R = CH₃) and the sum of B_{1a} and B_{1b} (R = C₂H₅) is not less than 97.0%. B_{1a}, and not less than 97.0% of 22,23-dihydroavermectin B₁ (B_{1a} + B_{1b}).

treatment assessment for loiasis and careful post-treatment follow-up should be implemented. The aim of this brief review is to assess whether any of these events could be attributed to the pharmacology of ivermectin, notably its affinity for the P-glycoprotein drug transporter, which might serve to reduce oral bioavailability of ivermectin and prevent its uptake into the brain, thereby preventing potentially fatal neurotoxicity.

Chemistry and Pharmacology of the Macrocyclic Lactones

The macrocyclic lactones comprise the avermectins (e.g. doramectin and ivermectin; [Figure 1]) and the milbemycins (e.g. moxidectin). They are natural fermentation products of *Streptomyces* bacteria. Many have potent anthelmintic and insecticidal properties [3,4] and are drugs of choice for nematode infections in animals. Ivermectin is the preferred treatment for onchocerciasis and other human filarial infections, such as *Wuchereria bancrofti*. Macrocyclic lactones produce a flaccid paralysis of the somatic worm musculature and inhibit feeding of the parasite through blockade of pharyngeal pumping [5–7], suggesting that a disruption of ingestion is the primary action of these agents [8]. However, the somatic musculature is also understood to be a target and may explain the

reduction in release of uterine microfilariae seen after exposure to ivermectin. Molecular genetics with *C. elegans* and expressed receptors has shown that macrocyclic lactones act as agonists of a family of invertebrate-specific inhibitory chloride channels that are activated by glutamic acid [8–11] and related phylogenetically to vertebrate GABA_A gated chloride channels [12]. While the selective effect of ivermectin can be explained by its action on the glutamate-gated chloride channels unique to invertebrates, at higher concentrations ivermectin can also potentiate vertebrate GABA_A gated chloride channels. This has led to suggestions that ivermectin and related drugs may be toxic in vertebrates having a deficiency in their blood brain barrier [13]. Recent findings suggest that the severe central nervous system side effects seen in various vertebrates following ivermectin treatment may be due to an absence of, or functional deficiency of P-glycoprotein [14].

The Blood Brain Barrier and p-glycoprotein

The blood brain barrier is comprised of brain capillary endothelial cells linked by tight junctions that form a lipophilic physical barrier that limits the passive transport of substances into the brain. While the permeability of the

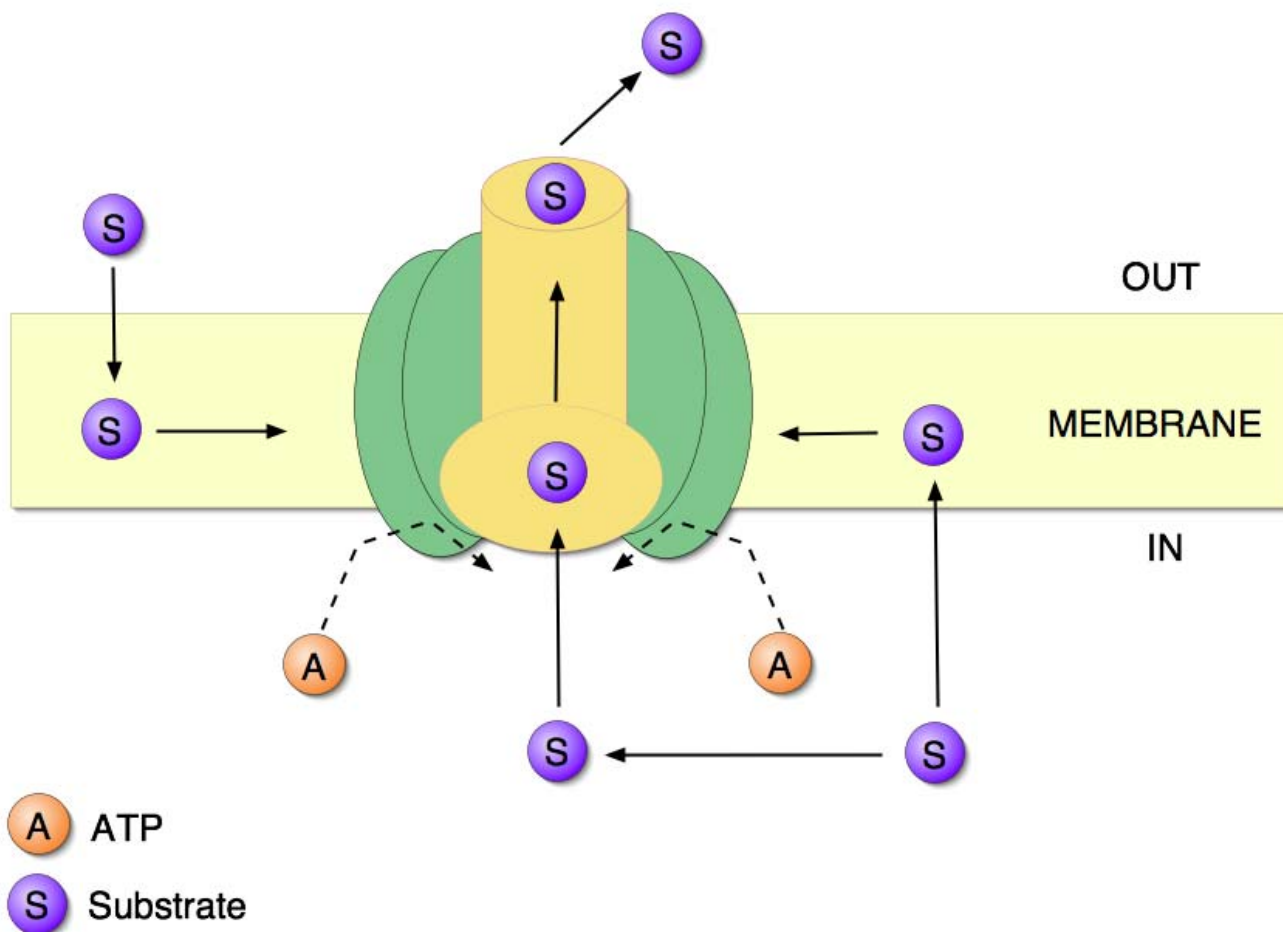


Figure 2

Simplified cartoon of P-glycoprotein structure and function: The P-glycoprotein molecule spans the cell membrane and in this way is in contact not only with the membrane but also the inside and the outside of the cell. The central portion of the molecule is a channel or pore through which toxic chemicals are pumped back out into the environment. The toxic chemicals can enter the transport pore either from the interior of the cell or from its membrane as shown. Molecules of ATP power the pumping action.

blood brain barrier increases with increasing lipophilicity, several transport proteins have been identified that regulate the penetration of many poorly lipophilic compounds. Conversely, many highly lipophilic substances such as cyclosporin and ivermectin show unexpectedly poor penetration of the blood brain barrier. It is now believed that this phenomenon is a result of the actions of drug efflux transporters [15,16]. Although molecular characterisation of those transport proteins present in the apical and basolateral membrane of brain capillary epithelial cells has identified a range of drug transporters such as MDR, MRP and OATP [17-20], it is

p-glycoprotein that has been most widely studied. P-glycoprotein is expressed in the apical membrane of brain capillary epithelial cells and is orientated to pump noxious substrates from inside cells and back into the blood (Figure 2). P-glycoprotein is responsible for limiting the brain penetration of a range of compounds of different therapeutic classes and in some cases may influence decisions regarding the clinical usage of such agents. For example, the anti-diarrhoeal opiate loperamide is safe and effective peripherally but does not act centrally since brain penetration is minimised through the actions of p-glycoprotein. However, it must be conceded that an accurate

assessment of the role of p-glycoprotein in the function of the human blood brain barrier is limited by our inability to measure readily drug concentrations in the brain. Furthermore, altered distribution within the brain may not always show itself as a change in circulating plasma concentrations. [16]. A pharmacodynamic indicator would be necessary to determine whether there was any exacerbation of centrally mediated effects brought about through genetic polymorphism in *mdr1* [21,22] or inhibition of p-glycoprotein function.

Neurotoxicology of Ivermectin

The principal toxicological effects of ivermectin seen in rats are ataxia, ptosis and decreased activity, and in dogs are mydriasis, tremors, ataxia and anorexia [23] with enhanced sensitivity in infant rats that have a poorly developed blood brain barrier (with regard to their deficiency in p-glycoprotein), and show higher brain plasma drug concentration ratios than adult rats. During the development of Mectizan® two reports, one from the veterinary use of ivermectin and another from a pre-clinical toxicology study, influenced the clinical development of Mectizan® such that a more conservative approach was taken [24]. First, it was noticed that a sub-population of collie dogs was remarkably sensitive to ivermectin-induced neurotoxicity [25]. This occurs at doses that are 1/200th of the dose required to cause toxicity in other dogs. Neurological manifestations of ivermectin in susceptible dogs include, hypersalivation, ataxia, blindness, coma, respiratory compromise, and death. It is now known that such ivermectin-sensitive collies exhibit a 4-bp deletion mutation of the *mdr1* gene that results in a frame shift, generating several stop codons that prematurely terminate synthesis of P-glycoprotein. Dogs that are homozygous for the mutation display the ivermectin sensitive phenotype, whereas those homozygous normal or heterozygous show no increased sensitivity to ivermectin [26]. Second, sub populations of CF-1 mice were identified as expressing reduced levels of P-glycoprotein, including a population that does not produce P-glycoprotein, identified as (-/-) [27]. Animals with the wild type (+/+) or deficient genotypes differ markedly in their sensitivity to ivermectin neurotoxicity and teratology, attributable to differential accumulation of ivermectin in brain and foetus. CF1 p-glycoprotein deficient mice are phenotypically identical to *mdr1a* and *mdr1b* knockout strains established in the laboratory [28]. Using such strains, differences in pharmacokinetics and tissue distribution, particularly accumulation in the brain, of several substances can be demonstrated when compared with p-glycoprotein wild type mice [29,30]. These authors clearly established the *mdr1a* transporter as central to limiting the oral bioavailability and brain uptake of these agents. Studies with ivermectin and the p-glycoprotein substrate cyclosporin A in CF-1 mice demonstrated enhanced absorption of iver-

mectin and cyclosporin A in the (-/-) strain although there were no differences in the intravenous pharmacokinetics of either drug. Hepatic drug metabolising capacity of (-/-) and (+/+) animals was similar. Concentrations of [³H]-ivermectin and [³H]-cyclosporin A were always higher in the brains of (-/-) mice after oral or intravenous administration but liver concentrations were identical to wild-type mice. These findings clearly demonstrate that changes in ivermectin disposition observed in the (-/-) mice arise through a deficiency in p-glycoprotein rather than any alteration in drug metabolism [14].

Drug-drug Interactions and Neurotoxicity of Ivermectin

Drug-drug interactions of clinical importance can occur when the pharmacology of a drug is altered by co-administration of another agent. The question of drug-drug interactions among anthelmintics is yet to be answered satisfactorily. This is an important issue, since combination chemotherapy is becoming more widespread as resistance to antiparasitic agents increases. For example albendazole and ivermectin are co-administered in programmes to eliminate lymphatic filariasis [31]. Experiments in human liver microsomes and recombinant enzyme systems have shown that ivermectin and albendazole are both substrates of cytochrome P450 (CYP) 3A4, the most widely distributed human P-450, which is expressed both in the liver and small intestine [32,33]. We have shown that albendazole is converted to albendazole sulphoxide, the major plasma metabolite in man, by microsomes prepared from human intestinal tissue [34]. There is, therefore, the potential for drug-drug interactions at each of these sites through competition for drug metabolising enzymes. The traditional view is that drug-drug interactions are a result of alterations in drug clearance mediated through a change in hepatic drug metabolism. However, it is becoming clear that some of these changes may arise through competition for binding sites on transport proteins. Fluorescent-labelled ivermectin (BOD-IPY-ivermectin) is extruded out of brain capillaries by a concentrative mechanism and this export process is reduced by substrates of p-glycoprotein without any change in drug uptake [35]. Moreover, transport of BOD-IPY-ivermectin is restricted to p-glycoprotein, as it is unaffected by specific inhibitors of related transport proteins and it interacts specifically with p-glycoprotein in functional kidney tubules [36]. These findings suggest that other P-glycoprotein substrates could compete with ivermectin at this site, resulting in reduced extracellular efflux and enhanced CNS toxicity. Such compounds include cyclosporin and HIV protease inhibitors [37,38]. Increased ivermectin concentrations accompanied by enhanced neurotoxicity of ivermectin in mice has already been demonstrated in the presence of cyclosporin [39]. While an interaction between ivermectin and albendazole could be postulated on the basis of these drugs being co-

substrates of CY3A4, it is not established that there could be an interaction at the level of p-glycoprotein. Although ivermectin is undeniably a substrate for P-glycoprotein, there is conflicting evidence for albendazole. Intestinal metabolism and secretion of albendazole sulphoxide into the intestinal lumen has been demonstrated and p-glycoprotein is reported to be involved in modulating resistance to albendazole in helminths [40,41]. However, experiments in Caco-2 cells failed to identify albendazole as a p-glycoprotein substrate or an inhibitor of p-glycoprotein mediated transport of digoxin [42], suggesting that a combination of the two drugs would not produce an adverse drug reaction based upon an interaction with p-glycoprotein. Albendazole sulphoxide or sulphone were not investigated.

Conclusions

P-glycoprotein is an integral component of the human blood brain barrier and plays a central role in limiting drug uptake into the brain. Altered expression or function of p-glycoprotein could conceivably allow elevation of brain concentrations of ivermectin and produce severe neurotoxicity. This might arise through a genetic polymorphism in p-glycoprotein or co-administration of ivermectin with a drug or foodstuff that might inhibit this efflux transporter. There are relatively few examples of clinically important drug-drug interactions at the blood-brain barrier that involve increased drug exposure within the brain. As mentioned previously, alterations in uptake into the brain may not be manifest as a change in systemic pharmacokinetics and may only become apparent when a pharmacodynamic assessment is made [16]. Furthermore, it may not be possible, with therapeutic doses, to achieve sufficiently high concentrations of a co-substrate to allow a drug-drug interaction to take place. For example, a range of antipsychotic, antiemetic and Ca²⁺ blocking agents failed to produce effective inhibition of p-glycoprotein despite the achievement of suitable concentrations within the systemic circulation [43]. Perhaps more importantly, it should be remembered that clinical and laboratory findings suggest that Mectizan[®]-associated *L. loa* encephalopathy is consistent with an embolic process triggered by massive microfilarial death. It may also involve circulating immune complexes or polymorphic inflammatory responses. Moreover, serious or even fatal encephalopathy may develop spontaneously, or following treatment with other microfilaricides such as diethylcarbamazine [44]. It is therefore questionable whether any of the neurological sequelae associated with administration of ivermectin are directly related to the drug and therefore it seems unlikely that any alterations in the pharmacokinetics of ivermectin for whatever reason would result in a more severe adverse reaction.

Competing Interests

None declared

Author's Contribution

Geoffrey Edwards was sole author.

Acknowledgements

Original work referred to by the author (references 32 and 34) was supported by a postgraduate studentship from GlaxoSmithKline Pharmaceuticals to Dr Helen Rawden.

References

- Gardon J, Gardon-Wendel N, Demanga-Ngangué, Kamgno J, Chippaux JP and Boussinesq M: **Serious reactions after mass treatment of onchocerciasis with ivermectin in an area endemic for *Loa loa* infection.** *Lancet* 1997, **350**:18-22.
- Boussinesq M, Gardon J, Gardon-Wendel N, Kamgno J, Ngoumou P and Chippaux JP: **Three probable cases of *Loa loa* encephalopathy following ivermectin treatment for onchocerciasis.** *Am J Trop Med Hyg* 1998, **58**:461-469.
- Burg RW and Stapley EO: **Isolation and characterisation of the producing organism.** In: *Ivermectin and Abamectin* Edited by: William C Campbell. New York, Berlin, Heidelberg, London, Paris, Tokyo, Springer-Verlag; 1989:24-32.
- Fisher MH and Mrozik H: **Chemistry.** In: *Ivermectin and Abamectin* Edited by: William C Campbell. New York, Berlin, Heidelberg, London, Paris, Tokyo, Springer-Verlag; 1989:1-23.
- Geary TG, Sims SM, Thomas EM, Vanover L, Davis JP, Winterrowd CA, Klein RD, Ho NF and Thompson DP: **Haemonchus contortus: ivermectin-induced paralysis of the pharynx.** *Exp Parasitol* 1993, **77**:88-96.
- Martin RJ: **An electrophysiological preparation of pharyngeal muscle reveals a glutamate-gated chloride channel sensitive to the avermectin analogue milbemycin D.** *Parasitology* 1996, **120**:587-594.
- Kotze AC: **Effects of macrocyclic lactones on ingestion in susceptible and resistant *Haemonchus contortus* larvae.** *J Parasitol* 1998, **84**:E631-635.
- Sangster NC and Gill J: **Pharmacology of anthelmintic resistance.** *Parasitol Today* 1999, **15**:141-146.
- Cully DF, Wilkinson H, Vassilatis DK, Etter A and Arena JP: **Molecular biology and electrophysiology of glutamate-gated chloride channels of invertebrates.** *Parasitology* 1996, **113**:191-200.
- Dent JA, Davis MW and Avery L: **avr-15 encodes a chloride channel subunit that mediates inhibitory glutaminergic neurotransmission and ivermectin sensitivity in *Caenorhabditis elegans*.** *EMBO J* 1997, **16**:5867-5879.
- Vassilatis DK, Arena JP, Plasterk RHA, Wilkinson HA, Schaeffer JM, Cully DF and Van der Ploeg LHT: **Genetic and biochemical evidence for a novel avermectin sensitive chloride channel in *Caenorhabditis elegans*.** *J Biol Chem* 1997, **272**:33167-33174.
- Kohler P: **The biochemical basis of anthelmintic resistance.** *Int J Parasitol* 2001, **31**:336-345.
- Etter A, Cully DF, Liu KK, Reiss B, Vassilatis DK, Schaeffer JM and Arena JP: **Picrotoxin blockade of invertebrate glutamate gated chloride channels: subunit dependence and evidence for binding within the pore.** *J Neurochem* 1999, **72**:318-326.
- Kwei GY, Alvaro RF, Chen Q, Jenkins HJ, Hop CEAC, Keohane CA, Ly VT, Strauss JR, Wang RW and Wang Z *et al.*: **Disposition of ivermectin and cyclosporin A in CF-1 mice deficient in MDR1A p-glycoprotein.** *Drug Metab Dispos* 1999, **27**:581-587.
- Tamai I and Atsuiji A: **Transporter-mediated permeation of drugs across the blood brain barrier.** *J Pharm Sci* 2000, **89**:1371-1388.
- Ayrton A and Morgan P: **Role of transport proteins in drug absorption, distribution and excretion.** *Xenobiotica* 2001, **31**:469-497.
- Cordon-Cardo C, O'Brien JP, Casals D, Rittman Graver L, Beidler JL, Melamed MR and Bertino JR: **Multidrug resistance gene P-glycoprotein is expressed by endothelial cells at blood-brain barrier sites.** *Proc Natl Acad Sci USA* 1989, **86**:695-698.

18. Huai-Yun H, Secrest DT, Mark KS, Carney D, Brandquist C, Elmquist WF and Miller DV: **Expression of multidrug resistance-associated protein (MRP) in brain microvessel endothelial cells.** *Biochem Biophys Res Comm* 1998, **243**:816-820.
19. Kusuvara H, Suzuki H, Naito M, Tsuro T and Sugiyama Y: **Characterisation of efflux transport of organic anions in a mouse brain capillary endothelial cell line.** *J Pharm Exp Ther* 1998, **285**:1260-1265.
20. Gao B, Steiger B, Noe B, Fritschy JM and Meier PJ: **Localisation of the organic anion transporting polypeptide 2 (Oatp 2) in capillary endothelium and choroids plexus epithelium of rat brain.** *J Histochem Cytochem* 1999, **47**:1255-1263.
21. Hoffmeyer S, Burk O, von Richter O, Arnold HP, Brockmoller J, John A, Cascorbi L, Gerlo T, Roots I, Eichelbaum M and Brinkmann U: **Functional polymorphisms of the human multidrug-resistance gene: multiple sequence variations and correlation of one allele with P-glycoprotein expression and activity in vivo.** *Proc Natl Acad Sci USA* 2000, **97**:3473-3478.
22. Kim RB, Leake B, Choo E, Dresser GK, Kubra SV, Schwarz UL, Taylor A, Xie HG, Stein CM and Wood AJJ et al.: **Identification of functionally important MDR1 variant alleles among African-American and Caucasian subjects.** *Drug Metab Rev* 2000, **32**(suppl 2):199.
23. Lankas GR and Gordon LR: **Toxicology.** In: *Ivermectin and Abamectin* Edited by: William C Campbell. New York, Berlin, Heidelberg, London, Paris, Tokyo, Springer-Verlag; 1989:89-112.
24. Brown KR: **Changes in the use profile of Mectizan®: 1987-1997.** *Ann Trop Med Parasitol* 1998, **92**:S61-S64.
25. Paul AJ, Tranquilli WJ, Seward RL, Todd KS Jr and DiPietro JA: **Clinical observations in colliers given ivermectin orally.** *Am J Vet Res* 1987, **48**:684-685.
26. Mealey KL, Bentjen SA, Gay JM and Cantor GH: **Ivermectin sensitivity in colliers is associated with a deletion mutation of the mdr1 gene.** *Pharmacogenetics* 2001, **11**:727-733.
27. Umbenhauer DR, Lankas GR, Pippert TR, Wise LD, Cartwright ME, Hall SJ and Beare CM: **Identification of a P-glycoprotein-deficient subpopulation in the CF-1 mouse strain using a restriction fragment length polymorphism.** *Toxicol Applied Pharmacol* 1997, **146**:88-94.
28. Schinkel AH, Smit JJ, van Tellingen O, Beijnen JH, Wagenaar E, van Deemter L, Mol CAAM, van der Valk MA, Robanus Maandag EC and Borst F: **Disruption of the mouse mdr1a P-glycoprotein gene leads to a deficiency in the blood-brain barrier and to increased sensitivity to drugs.** *Cell* 1994, **77**:491-502.
29. Schinkel AH, Wagenaar E, van Deemter L, Mol CAAM and Borst F: **Absence of the mdr1a P-glycoprotein in mice affects tissue distribution and pharmacokinetics of dexamethasone, digoxin and cyclosporin A.** *J Clin Invest* 1995, **96**:1698-1705.
30. Sparreboom A, van Asperen J, Mayer U, Schinkel AH, Smit JW, Meier DKF, Borst P, Nooijen WJ, Beijnen JH and van Tellingen O: **Limited oral bioavailability and active epithelial secretion of paclitaxel (taxol) caused by P-glycoprotein in the intestine.** *Proc Natl Acad Sci USA* 1997, **94**:2013-2035.
31. Horton J, Witt C, Ottesen EA, Lazdins JK, Addiss DG, Awadzi K, Beach MJ, Belizario YY, Duno SK and Espinel M et al.: **An analysis of the safety of the single dose, two drug regimens used in programmes to eliminate lymphatic filariasis.** *Parasitology* 2000, **121**: Suppl:S147-160.
32. Rawden HC, Kokwaro GO, Ward SA and Edwards G: **Relative contribution of cytochromes P-450 and flavin-containing monooxygenases to the metabolism of albendazole by human liver microsomes.** *Br J Clin Pharmacol* 2000, **49**:313-322.
33. Zeng, Andrew NW, Arison BH, Luffer-Atlas D and Wang RW: **Identification of P4503A4 as the major enzyme responsible for the metabolism of ivermectin by human liver microsomes.** *Xenobiotica* 1998, **28**:313-321.
34. Rawden HC: **Ph.D. thesis. "An investigation of potential chemotherapeutic drug combinations with Albendazole for the treatment of Echinococcus infection".** *The University of Liverpool* 1999.
35. Nobmann S, Bauer B and Fricker G: **Ivermectin excretion by isolated functionally intact brain endothelial capillaries.** *Br J Pharmacol* 2001, **132**:722-728.
36. Fricker G, Gutmann H, Droulle A, Drewe J and Miller DS: **Epithelial transport of anthelmintic ivermectin in a novel model of isolated proximal kidney tubules.** *Pharm Res* 1999, **16**:1570-1575.
37. Drewe J, Gutmann H, Fricker G, Torok M, Beglinger C and Huwyler J: **HIV protease inhibitor ritonavir: a more potent inhibitor of P-glycoprotein than the cyclosporine analog SDZ PSC 833.** *Biochem Pharmacol* 1999, **15**:1147-1152.
38. Gutmann H, Fricker G, Drewe J, Toeroek M and Miller DS: **Interactions of HIV protease inhibitors with ATP-dependent drug export proteins.** *Mol Pharm* 1999, **56**:383-389.
39. Marques-Santos LF, Bernardo RR, de Paula EF and Rumjanek VM: **Cyclosporin A and trifluoperazine, two resistance modulating agents, increase ivermectin neurotoxicity in mice.** *Pharmacol Toxicol* 1999, **84**:125-129.
40. Nare B, Liu Z, Prichard RK and Georges EE: **Benzimidazoles, potent anti-mitotic drugs: substrates for the P-glycoprotein transporter in multidrug-resistant cells.** *Biochem Pharmacol* 1994, **48**:2215-2222.
41. Redondo PA, Alvarez AI, Garcia JL, Larrode OM, Merino G and Prieto JG: **Presystemic metabolism of albendazole: experimental evidence of an efflux process of albendazole sulphoxide to intestinal lumen.** *Drug Metab Dispos* 1999, **27**:736-740.
42. Merino G, Alvarez AJ, Prieto JG and Kim RB: **The anthelmintic agent albendazole does not interact with p-glycoprotein.** *Drug Metab Dispos* 2002, **30**:365-369.
43. Ibrahim S, Peggins J, Knapton A, Licht T and Aszalos A: **Influence of antipsychotic, antiemetic and Ca²⁺ channel blocker drugs on the cellular accumulation of the anticancer drug daunorubicin: P-glycoprotein modulation.** *J Pharm Exp Ther* 2000, **295**:1276-1283.
44. Carme B, Boulesteix J, Boutes H and Puruehnce MF: **Five cases of encephalitis during treatment of loiasis with diethylcarbamazine.** *Am J Trop Med Hyg* 1991, **44**:684-690.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:
http://www.biomedcentral.com/info/publishing_adv.asp

